



EVALUATION OF BIO-INTENSIVE DISEASE MANAGEMENT OF FUSARIUM ROT IN SMALL CARDAMOM (BIDMFC) WITH LOCALLY ISOLATED STRAINS

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Abstract: The cultivation of small cardamom and maintenance of healthy plants have become difficult tasks due to the incidence and spread of Fusarium infections in recent years. Fusarium infection in small cardamom have been reported in the form of capsule infection in the field, seed rot and seedling wilt in nurseries, stem rot & stem lodging in plantations, rhizome rot, root tip rot and foliar yellowing. Field experiment was conducted at Santhanpara, Vandannedu, Parathode and Kattappana to evaluate the individual and combined efficacy of *Pseudomonas fluorescens* (IDK-S-1, IDK-V-2, IDK-P-3, and IDK-k-4), *Bacillus subtilis* (IDK-S-1, IDK-V-2, IDK-P-3, and IDK-k-4), *Trichoderma harzianum* (IDK-S-1, IDK-V-2, IDK-P-3, and IDK-K-4) and AMF (IDK-S-1, IDK-V-2, IDK-P-3, and IDK-k-4) strains to promote the growth and yield parameters of small cardamom and to manage Fusarium wilt disease in field conditions. The dominant pathogen which causes Fusarium wilt of small cardamom was isolated and identified as *Fusarium oxysporum* Schlecht. Five native bacterial antagonists and fungus were isolated from forest and healthy small cardamom plantation soil in different geographical regions. Under in field conditions, the results revealed that the foliar spray and drenching of combined application of PF (Pf-IDK-S-1)+BS(Bs-IDK-V-2) was found to effectively inhibit the mycelial growth of the pathogen (by 60%) when compared to application of individual strains of the bacterial antagonists. The above strains of *P. fluorescens* and *B. subtilis* (PF (Pf-IDK-S-1) +BS (Bs-IDK-V-2)) were found compatible. The soil combined application of PF (IDK-S-1) +TH (IDK-P-3) exhibited the highest disease reduction. Also, small cardamom plantation treated with PF (IDK-S-1) + BS (IDK-V-2) strains showed a significant stimulatory effect on plant height and increased the yield up to 27% in comparison to the non-bacterized control. The combined strains also increased small cardamom capsule fruit weight. It could be concluded that synergistic consortia of beneficial bacteria isolated from rhizosphere soil are perfectly able to promote plant growth and could be exploited for sustainable management of soil borne diseases especially, Fusarium rot of small cardamom.

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1. INTRODUCTION

Fusarium rot caused by *Fusarium oxysporum* Schlecht is one of the most widespread and important fungal diseases of small cardamom (Thomas and Vijayan, 2002). *F. oxysporum* disease may become destructive at all growth stages of the crop under favourable weather conditions and can cause severe infection in this changing climate scenario. Dhanya *et al.* (2018) reported 50 per cent yield loss from poorly managed plants due to the disease. The pathogen survives in soil and crop residues for many years making the management of the pathogen very difficult. The *Fusarium* sp. initially attacks the tiller producing pale discolored lesions leading to dry rotting. The

infected tillers are weakened at the point of attack resulting in partial breakage. These tillers bend down and hang from the point of infection (Josephraj kumar *et al.*, 2007). Symptoms include burnt appearance of panicles and rotting of roots from the tip causing yellowing of the plant and thereby reduction in the yield (Murugan *et al.*, 2016). In the scenario of growing concerns over environment pollution and health hazards the need for an integrated approach in the disease management strategy is of high importance. The integrated disease management using various bio-control agents and bio inputs improved the vegetative growth and the yield of the plant in addition to disease management of Fusarium rot of cardamom

(Dhanya *et al.*, 2018). The present study entitled as “Evaluation of Bio-Intensive Disease Management of Fusarium Rot in Small Cardamom (BIDDMFC) With

Locally Isolated Strains” was undertaken with an objective to assess the

Fusarium rot severity in Idukki district, develop a bio-intensive disease management practices for the disease and to study the impact of these practices on the soil and plant health.

is prevalent during the summer season, survey was carried out in February- May 2021-2023 in two major cardamom cultivated blocks of Idukki districts viz., Santhanpara and Vandanmedu. One Panchayat each of above blocks were surveyed. Three plantations of size 1ha each were selected from each panchayat and the details are given below. Each plantation was divided into four plots each with 250 plants. Ten plants were selected from each plot and five tillers randomly selected from each plant were scored. The following score chart developed by Dhanya *et al.* (2018) used for the study.

MATERIALS AND METHODS

The location of current study entitled Evaluation of Bio-Intensive Disease Management of Fusarium Rot in Small Cardamom (BIDDMFC) With Locally Isolated Strains” was Small Cardamom Plantation at Santhanpara, Vandanmedu, Parathode and Kattappana in Idukki during the period of 2021-2023. *Fusarium oxysporum* diseases infection in cardamom plantation

0= No disease

1= 1-10 % of tillers or panicles had fungal lesions 2= 11-25 % of tillers or panicle had fungal lesions

3=26-50 % of tillers had fungal lesions with drying of panicles from tip 4= 51-75 % of tiller stake hold of fungal lesions with partial panicle blight

Based on the data, disease severity (PDI) was calculated using the formula

$$\text{PDI} = \frac{\text{Sum of score}}{\text{observed maximum score given}} \times \frac{100}{\text{Total number of tillers}}$$

Percent Disease Index was calculated using the formula of Singh (2002).

$$\text{Disease Incidence} = \frac{\text{No. of infected plants}}{\text{Total number of plants observed}} \times 100$$

Root staining for Arbuscular Mycorrhizal Fungi:

At the end of the experiment to study the mycorrhizal colonization pattern of AMF inoculated the plant roots were collected, and washed with water. 10 % KOH (W/V) was added to container and made root samples were submerged but the fluid did not fill more than the half of the container. The tissues were soaked overnight and replaced with fresh KOH before the heat treatment. The solution turned into brown

colour by overnight that indicated the tannins were left from tissues. Heat treatment was given by boiling the processed roots in a water bath. The sample solution was removed and rinsed with tap water for three times. After that 5% HCl was added and allowed to retain for about a minute. After that HCl was poured out and added trypan blue stain and examined the stained roots under a microscope (Philip and Haymann, 1970).

$$\text{Per cent root colonization} = \frac{\text{No. of root bits showing colonization}}{\text{Total number of root bits observed}} \times 100$$

Five year old well established plantations with yielding plants of most popular local variety Njallani, which is highly susceptible to the disease, was chosen for the survey. Different treatments of bio-control agents were scheduled into seven treatments as basal applications and foliar spraying. Field experiment was conducted at Santhanpara, Vandanmedu, Parathode and Kattappana to evaluate the individual and

combined efficacy of *Pseudomonas fluorescens* (IDK-S-1, IDK-V-2, IDK-P-3, and IDK-k-4), *Bacillus subtilis* (IDK-S-1, IDK-V-2, IDK-P-3, and IDK-k-4), *Trichoderma harzianum* (IDK-S-1, IDK-V-2, IDK-P-3, and IDK-K-4) and AMF (IDK-S-1, IDK-V-2, IDK-P-3, and IDK-k-4) strains to promote the growth and yield parameters of small cardamom and to manage Fusarium wilt disease in field conditions.

Table 1. Treatment details of different local strains to manage Fusarium rot of small cardamom

Treatments	*IDK-S-1	*IDK-V-2	*IDK-P-3	*IDK-k-4
T1- <i>Pseudomonas fluorescens</i>	Spraying of PF- IDK-S-1 @2.5 ml per L of water (3 times)	Spraying of PF- IDK-V-2 2.5 ml per L of water (3 times)	Spraying of PF- IDK-P-3 2.5 ml per L of water (3 times)	Spraying of PF- IDK-k-4 2.5 ml per L of water (3 times)
T2- <i>Bacillus subtilis</i>	Spraying of BS- IDK-S-1 2.5 ml per L of water (3 times)	Spraying of BS- IDK-V-2 2.5 ml per L of water (3 times)	Spraying of BS- IDK-P-3 2.5 ml per L of water (3 times)	Spraying of BS- IDK-k-4 2.5 ml per L of water (3 times)
T3- <i>Trichoderma harzianum</i>	Basal application of TH-IDK-S-1 (1kg) with Neem cake (10kg) and FYM(90kg)	Basal application of TH-IDK-V-2 (1kg) with Neem cake (10kg) and FYM(90kg)	Basal application of TH-IDK-P-3 (1kg) with Neem cake (10kg) and FYM(90kg)	Basal application of TH-IDK-k-4 (1kg) with Neem cake (10kg) and FYM(90kg)
T4-AMF	Basal application of AMF -IDK-S-1 @ 50gm to the root zone at the time of planting (3 times)	Basal application of AMF IDK-V-2 @ 50gm to the root zone at the time of planting (3 times)	Basal application of AMF -IDK-P-3 @ 50gm to the root zone at the time of planting (3 times)	Basal application of AMF -IDK-k-4 @ 50gm to the root zone at the time of planting (3 times)
T5- <i>Pseudomonas fluorescens</i> + <i>Bacillus subtilis</i>	Spraying of PF+BS- IDK-S-1 @2.5 ml per L of water (3 times)	Spraying of PF+BS- IDK-V-2 2.5 ml per L of water (3 times)	Spraying of PF+BS- IDK-P-3 2.5 ml per L of water (3 times)	Spraying of PF- IDK-k-4 2.5 ml per L of water (3 times)
T6- <i>Trichoderma harzianum</i> + AMF	Basal application of TH+AMF -IDK-S-1 @ 50gm to the root zone at the time of planting	Basal application of TH+AMF IDK-V-2 @ 50gm to the root zone at the time of planting	Basal application of TH+AMF -IDK-P-3 @ 50gm to the root zone at the time of planting	Basal application of TH+AMF -IDK-k-4 @ 50gm to the root zone at the time of planting
T7 Control (untreated plants)	-	-	-	-

***IDK-S-1-Santhanpara Local Strains, IDK-V-2-Vandanmedu Local Strains, IDK-P-3-Parathode and IDK-K-4-Kattappana.**

Results and Discussion:

A field experiment was conducted to assess the efficacy of selected bio-agents (individually and in combination) for the management of the disease in Idukki district of Kerala in CRD using seven treatments with three replications. Basal application of TH+AMF -IDK-S-1, TH+AMF IDK-V-2, TH+AMF -IDK-P-3 and TH+AMF -IDK-k-4 @ 50gm with the individual of Trichoderma and AMF different strains and combination of TH+AMF to the root zone at the

time of planting, Spraying of PF+BS-IDK-S-1, PF+BS-IDK-V-2, PF+BS-IDK-P-3, PF- IDK-k-4 @2.5 ml per L of water with individual and combination of bio-agents at monthly interval for three times resulted in average effective disease management (disease incidence: 30% and disease severity: 24.26%) compared to the inoculated control (disease incidence: 90% and disease severity: 59.38 %) (Table.2 & 3).

Table.2. Biometric characters and Disease incidence and severity of cardamom plants inoculated with *Fusarium sp.* in response to treatments

Treatment	Plant Height (cm)	No. of Tillers	No. of Leaves	Leaf length(cm)	Leaf width (cm)	Disease incidence	Disease severity
T1	48.05	3.0	13	38	10.00	78.83(84.00) ^e	36.23(40.00) ^{cd}
T2	64.10	3.0	13	37	12.50	46.43(52.50) ^c	32.89(30.00) ^{bc}
T3	69.30	3.0	12	46	10.50	37.81(41.00) ^b	29.67(19.50) ^a
T4	75.70	3.0	14	59	11.25	41.56(47.50) ^c	36.60(27.50) ^{bc}
T5	92.00	4.0	17	63	13.66	59.25(65.66) ^d	39.75(34.50) ^{bcd}
T6	96.50	4.0	15	62	12.83	36.27(34.33) ^a	27.26(19.00) ^a
T7	40.45	2.0	9	41	7.80	84.63(96.50) ^d	65.23(80.00) ^{cd}

In the present study, about four strains of PGPR were isolated from the different location of small cardamom plantation. The field trials were clearly showed that the bacterial antagonists of *Pseudomonas fluorescens* (IDK-S-1-Santhanpara Local Strains, IDK-V-2-Vandanmedu Local Strains, IDK-P-3-Parathode and

IDK-K-4-Kattappana) and *B. Subtilis* strains (IDK-S-1-Santhanpara Local Strains, IDK-V-2-Vandanmedu Local Strains, IDK-P-3-Parathode, IDK-K-4-Kattappana) were compatible and effectively inhibited the growth of *Fusarium* disease in small cardamom plantation.

Table 3. Different Local strains of Biometric characters and Disease incidence and severity of cardamom plants inoculated with *Fusarium sp.* in response to treatments

LS	Plant Height (cm)	No. of Tillers	No. of Leaves	Leaf length (cm)	Leaf width (cm)	DS	DS	RC
IDK-S-1	65.10	3.0	11	46	10.5	42.20	31	65.6
IDK-V-2	77.10	4.0	19	59	11.2	39.27	27	78.2
IDK-P-3	82.00	3.0	18	53	13.2	36.27	29	73.5
IDK-K-4	86.00	4.0	16	52	12.1	56.45	32	69.2
Control	43.45	2.0	8	31	9.6	84.34	65.38	0.00

The aim of the present work is to ascertain that small cardamom plantation increase productivity when AMF (IDK-S-1-Santhanpara Local Strains, IDK-V-2-Vandanmedu Local Strains, IDK-P-3-Parathode and IDK-K-4-Kattappana) and Trichoderma (IDK-S-1-Santhanpara Local Strains, IDK-V-2-Vandanmedu Local Strains, IDK-P-3-Parathode and IDK-K-4-Kattappana) are combined applied, and how such an effect can be ruled. This simultaneous application of a *Trichoderma harzianum* bio-control strain and an AMF formulation produces a significant increase in the colonization by Trichoderma and the presence of AMF in Small cardamom plantation. Expression profiling of defense-related marker genes suggests that the phytohormone salicylic acid plays a key role in the modulation of the root colonization process when both fungi are jointly applied. As a conclusion drawn from

the data on vegetative characters revealed that application of *Bacillus subtilis* and *P. fluorescens* as single basal application and in combination with *P. fluorescens* spray (T5 and T6) showed maximum plant height and leaf length and number of leaves. Sole application of *P. fluorescens* also promoted plant height and number of leaves. There for the treatments comprised of *P. fluorescens* and *Bacillus subtilis* individually and in combination resulted in good vegetative growth of treatment plants.

CONCLUSION:

The present study demonstrated that combined application of bacterial and Fungus native antagonistic (IDK-S-1-Santhanpara Local Strains, IDK-V-2-Vandanmedu Local Strains, IDK-P-3-Parathode and IDK-K-4-Kattappana) is a promising approach for the

eco friendly management of Fusarium wilt disease caused by *F. oxysporum* enhancing the growth of the Small cardamom plantation.

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References

- [1]. Amini, J. and Sidovich, D.F. 2010. The effects of fungicides on *Fusarium oxysporum* f. sp. *Lycopersici* associated with *Fusarium* wilt of tomato. *J. Plant Protec. Res.* 50(2): 172- 178.
- [2]. Anguilar, E.A., Turner, D.W. and Sivasithamparam, K. 2000. *Fusarium oxysporum* f. sp. *cubense* inoculation and hypoxia alter peroxidase and phenylalanine ammonia lyase activities in nodal roots of banana cultivars (*Musa* sp.) differing in their susceptibility to *Fusarium* wilt. *Aust. J. Bot.* 48: 589–596.
- [3]. Animisha., Zachariya, S., Jaisal, K.K. and Pandey, P. 2012. Integrated management of chickpea wilt incited by *Fusarium oxysporum* f. sp. *ciceris*. *Int. J. Agric. Res.* 7(5): 284-290.
- [4]. AOAC (Association of Official Analytical Chemist) 1995. *Official methods of analysis*: 15th Ed.), Washington, DC, 769p.
- [5]. Arfaoui, A., El Hadrami, A. Mabrouk, Y., Sifi, B., Boudabous, A., El Hadrami, I., Daayf, F. and Cherif, M. 2007. Treatment of chickpea with *Rhizobium* isolates enhances expression of phenylpropanoid defense-related genes in response to infection by *Fusarium oxysporum* f. sp. *ciceris*. *Plant Physiol. Biochem.* 45: 470-479.
- [6]. Audipudi, A. V., Pradeepkumar, N., and Allu, S. 2016. Effect of mixed inoculations of plant growth promoting rhizobacteria of chilli on growth and induced systemic resistance of *Capsicum frutescens* L. In: Sayyed, R.Z., Reddy, M.S., and AL-Turki, A.I. (ed.), *Recent Trends In PGPR Research For Sustainable Crop Productivity*. Scientific Publishers, Delhi, pp. 9-20.
- [7]. Azarmi, R. Hajieghrari, B., and Giglou, A. 2011. Effect of *Trichoderma* isolates on tomato seedling growth response and nutrient uptake. *Afr. J. Biotechnol.* 10(31): 5850-5855.
- [8]. Balliu, A., Sallaku, G., and Rewald. 2015. AMF inoculation enhances growth and improves the nutrient uptake rates of transplanted, salt-stressed tomato seedlings. *sustainability* 7: 15967-15981.
- [9]. Bardia, P.K. and Rai, P.K. 2007. *In vitro* and field evaluation of biocontrol agents and fungicides against wilt of cumin caused by *Fusarium oxysporum* f. sp. *cuminii*. *J. Spices Aromat. Crops* 16(2) : 88–92.
- [10]. Basu, M.J. and Santhaguru, K. 2009. Impact of *Glomus fasciculatum* and fluorescent pseudomonads on growth performance of *Vigna radiate* (L.) Wilczek challenged with phytopathogens. *J. Plant Prot. Res.* 49:190-194.
- [11]. Bender, S. F., Conen, F., Marcel, G.A., and der Heijden, V. 2015. Mycorrhizal effects on nutrient cycling, nutrient leaching and N₂O production in experimental grassland. *Soil Biol. Biochem.* 80: 283-292.
- [12]. Bisht, R., Chaturvedi, S., Srivastava, R., Sharma, A.K., and Johri, B.N. 2009. Effect of arbuscular mycorrhizal fungi, *Pseudomonas fluorescens* and *Rhizobium leguminosarum* on the growth and nutrient status of *Dalbergia sissoo* Roxb. *Trop. Ecol.* 60(2): 231-242.
- [13]. Boukerma, L., Benchabane, M., Charif, A., and Khelifi. 2017. Activity of plant growth promoting Rhizobacteria (PGPRs) in the biocontrol of tomato *Fusarium* wilt. *Plant Prot. Sci.* 53(2): 78-84.
- [14]. Bray, G.G. and Thrope, W.V. 1954. Analysis of phenolic compounds of interest in metabolism. *Methods Biochem. Anal.* 1:27-52.
- [15]. Brito, I., Goss, M.J., Alho, L., Brigido, C., van Tuinen, D., Felix, M.R. and Carvalho, M. 2018. Agronomic management of AMF functional diversity to overcome biotic and abiotic stresses- the role of plant sequence and intact extraradical mycelium. *Fungal Ecol.* 30: 1-10.
- [16]. Buiatti, M., Scala, A., Bettini, P., Nascari, G., Morpurgo, R., Bogani, P., Pellegrini, G., Gimelli, G. 1984. Correlations between *in vivo* resistance to *Fusarium* and *in vitro* response to fungal elicitors and toxic substances in carnation. *Theor. Appl. Genet.* 70:42-47.
- [17]. Cacique, I, S., Pinto, L.F.C.C., Aucique-Pierrez, C.E., Filho, J.A.W., and Rodrigues, F.A. 2020. Physiological and biochemical insights into the basal level of resistance of two maize hybrids in response to *Fusarium verticillioides* infection. *Plant Physiol. Biochem.* 152: 194-210.
- [18]. Carpio L. A., Davies F. T., Arnold M. A. 2005. Arbuscular mycorrhizal fungi, organic and inorganic controlled-release fertilizers: effect on growth and leachate of container-grown Carrasco, A., Boudet, A.M. and Marigo.

- G. 1978. Enhanced resistance of tomato plants to *Fusarium* by controlled stimulation of their natural phenolic production. *Physiol. Plant Pathol.* 12: 225-232.
- [19]. Cavagnaro, T.R., Bender, S.F., Asghari, H.R., and de Heijden, M.G.A. 2015. The role of arbuscular mycorrhizas in reducing soil nutrient loss. *Trends Plant Sci.* 20(5):284-190.
- [20]. Chang, M.M., Hadwiger, L.A. and Horovitz, D. 1992. Molecular characterization of a pea β -1,3-glucanase induced by *Fusarium solani* and chitosan challenge. *Plant Mol.Biol.* 20: 609-618.
- [21]. Chang, P.F.L., Hsu, C.C., Lin, Y.H., Chen, K.S., Huang, J.W. and Liou, T.D. 2008. Histopathology comparison and phenylalanine ammonia lyase (PAL) gene expressions in *Fusarium* wilt infected watermelons. *Aust. J. Agric. Res.* 59:1146-1155.
- [22]. Chang, T.H., Lin, Y.H., Chen, K.S., Huang, J.W., Hsiao, S.C. and Chang, P.F.L. 2015. Cell wall reinforcement in watermelon shoot base related to its resistance to *Fusarium* wilt caused by *Fusarium oxysporum* f. sp. *niveum*. *J. Agric. Sci.* 153: 296-305.
- [23]. Christopher, D.J., Raj, T.S., Rani, S.U., and Udayakumar, R. 2010. Role of defense enzymes activity in tomato as induced by *Trichoderma virens* against *Fusarium* wilt caused by *Fusarium oxysporum* f. sp. *lycopersici*. *J. Biopesticide* 3(1): 158-162.
- [24]. Colla, G., Roupheal, Y., Bonini, P., and Cararelli. 2015. Coating seeds with endophytic fungi enhances growth, nutrient uptake, yield and grain quality of winter wheat. *Int. J. Plant Prod.* 9(2): 1735-6814.
- [25]. Corsini, D.L. and Pavek, J.J. 1980. Phenylalanine ammonia lyase activity and fungitoxic metabolites produced by potato cultivars in response to *Fusarium* tuber rot. *Physiol. Plant Pathol.* 16: 63-72.
- [26]. Curir, P., Dolci, M., Dolci, P., Lanzotti, V. and Dee Cooman, L. 2003. Fungitoxic Phenols from Carnation (*Dianthus caryophyllus*) Effective Against *Fusarium oxysporum* f. sp. *dianthi*. *Phytochem. Anal.* 14: 8-12.
- [27]. Datta, J. and Lal, N. 2018. Temporal and spatial changes in host defense enzymes in response to *Fusarium* Wilt in Pigeonpea (*Cajanus cajan* (L.) Millspaugh). *Int. J. Biochem.Res.Rev.* 23(1): 1-15.
- [28]. Dhanapal, K. and Thomas, J. 1996. Evaluation of *Trichoderma* isolates against rot pathogens of cardamom. In: Rao, K.M. and Mahadevan, A. (ed.). *Recent Advances in Biocontrol of Plant Pathogens*. Today and Tomorrow Printers and Publishers, New Delhi, India. pp. 67-75.
- [29]. Dhanya, M.K., Murugan, M., Deepthy, K.B., Aswathy, T.S. and Sathyan, T. 2018. Management of *Fusarium* rot in small cardamom. *Indian. J. Plant Prot.* 46(1): 57-62.

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