Abating of Bemisia tabaci (Gennadies) using Amblyseius cydnodactylon (Shehta &Zoher) in the same ecological niche (Inseta, Alevrodidae & Acari, phytoseiide)

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Abstract: The phytoseiide mite *Amblyseius cydnodactylon (Shehta and Zoher)* as predator on *Bemisia tabaci* (Gennadies) was studied at laboratory conditions, 25±2°C and 70±5% R.H. The predator was able to feed and complete its development on the tested prey. The larval stage developed to the protonymphal stage without feeding. On the other hand, the consumption rate increased through the developmental stages, respectively. Maximum mean for the total food consumption of the female predator was recorded during the oviposition period, it consumed an average of (242.61) preys, while the average number of *B. tabaci* devoured during life span were (303. 8 and 212.65) preys for female and male, respectively. This study declared that *A. cydnodactylon* may be considered of great value for controlling *B. tabaci* under greenhouses and field crops in Egypt.

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1. Introduction

Hemipteran pests can seriously damage crops by feeding directly on plant sap, excreting honeydew, and transmitting more than 110 plant viruses Jones, 2003, Mughra, et al., 2008 and El-Sharkawy 2013. The whitefly, Bemisia tabaci (Gennadius) (Hemiptera: Aleyrodidae), is a complex species, it firstly described in Greece by (Gennadius, 1889), it has become one of the most serious agricultural and ornamental plant pests in many areas of the world in recent decades (DeBarro, 1995; Tanigoshi, et al. 2001; Hodges and Evans 2005; Oiu, et al. 2007 and Oiu, et al., 2009). This pest completed its development on 118 species of plants in 79 genera belonging to 28 families and caused damage through feeding, causing sooty mold by its honey dew, transmitting more than 111species of plant pathogenic viruses, and inducing plant physiological discords (Gerling, 1990, Heinz 1996; Brown, 2001 and Abd-Rabou & Simmons 2010). Historically, B. tabaci has been difficult to control with conventional insecticides in agronomic and horticultural production systems (Palumbo et al., 2001).

The diet of phytoseiid mites may include several whitefly and mite species as well as pollen in several agro ecosystems. The whitefly, *B. tabaci*is a pest with high resistance to chemical insecticides that occur in greenhouses in temperate regions (El-Banhawy, *et al.*, 2000 and Nomikou, *et al.*, 2001).

The predacious mite, *Amblyseius cydnodactylon* Shehata and Zaher (Acari, Phytoseiidae) is a common natural enemy inhabiting low growing plants like cucumber in Egypt. It has been recorded associated

with infestations of the two-spotted spider mite, *Tetranychus urticae* Koch (Acari, Tetranychidae) and the whitefly, *Bemesia tabaci* (Genn.). The mite *A. cydnodactylon*is considered a generalist predator and it was recorded preying on several pests including tetranychid and eriophyid mites in addition to nymphs of whiteflies and thrips (**El-Banhawy** *et al.*, **2001**).

However, the present study was conducted to evaluate the effect of laboratory conditions on the developmental life stages, life cycle, longevity, fecundity, lifespan and prey consumption of *A. cydnodactylon* as a biocontrol agent controlling the insect pest, *B. tabaci*.

2. Material and Methods Prev culture:

The whitefly, *Bemisia tabaci* (Gennadius) was found and daily obtained from leaves of castor oil plants, *Ricinus communisi* L. (Euphorbiaceae) at Hehia district, El-Sharkia Governorate, Egypt .during 2014/2015 season.

Predator culture:

A laboratory colony of *Amblyseius cydnodactylon* Shehata and Zaher occurring with mulberry plant leaves, *Morus alba* L. (Moraceae) at the same area. Mass culture was maintained in the laboratory on castor leaves infested with *Tetranychus urticae Koch* (Acari, Tetranychidae) as prey.

Experimental procedure:

The experiment was conducted under the laboratory conditions, 25 ± 2 °C and $70\pm 5\%$ R. H. Thirty gravid females of *A. cydnodactylon* were taken randomly and transferred to rearing substrates. The

rearing substrates were disks of castor oil plants leaves (one inch in diameter), placed on cotton moistened with water in Petri-dishes (6 cm in diameter). Females were left 24 hours and their oviposited eggs are considered the first step of biological aspects. Thereafter, when a sufficient number of eggs were laid, the adult females were removed and thus eggs from the same age were obtained to start the investigation. Observations were made every 6 hours intervals to observe the eggs hatching. As the eggs hatched into larvae, individuals were transferred very carefully onto fresh leaf disks of castor leaves (one inch in diameter). Leaf discs were placed with the upper surface facing down on cotton layer, moistened with water in a Petri-dishes (6 cm in diameter). Water was added when needed to maintain the suitable moisture. The leaf margin was surrounded by tangle foot (candabalsm, castor beans & eitronella oils) to prevent the escaping of mites. Twenty replicates were maintained for the experiment. All Petri dishes were kept under laboratory conditions. Crawlers of B. tabaci were given as food for the predatory mite, A. cydnodactylon. The biological aspects for female and male predator were recorded by a stereomicroscope binocular.

Statistical analysis:

Data were statistically analyzed using the analysis of variance according to **Sendecor and Cochran (1982)** using the computer program **SPSS (2006).**

3. Results

Developmental stages of *Amblyseius* cydnodactylon Shehata and Zaher fed on crawlers of *B. tabaci* (Genn.) at laboratory conditions are presented in (Tables 1 & 2).

The incubation period for the eggs of *A. cydnodactylon* averaged 1.8 and 1.71 days for female and male, respectively. After hatching, the larval stage developed to the protonymphal stage without feeding, remained active for a short period of 1.41and 1.26 days for female and male, respectively. The duration period of the female and male protonymphs recorded 2.45 and 2.11 days after molting to give the deutonymphs. Generally, the immature stages and life cycle required average periods of (7.45 & 6.38) and (9.25 & 8.19) days for female and male, respectively (Table 1).

Table (1): Developmental stages of A. cydnodactylon, fed on crawlers of B. tabaci at 25±2 °C and 70±5% R.H..

Stages	Mean ± SE. days		
_	Female	Male	
Incubation period	1.8±0.258	1.71±0.441	
Active larva	1.41±0.137	1.26±0.473	
Protochrysalis	0.75±0.375	0.6±0.029	
Larval stage	2.1±0.394	1.91±0.013	
Active protonymph	1.68±0.544	1.31±0.118	
Deutochrysalis	0.81±0.044	0.77±0.353	
Protonymphal stage	2.45±0.438	2.11±011	
Active deutonymph	2.11±0.353	1.71±0.071	
Tritochrysalis	0.83±0.125	0.65±0.016	
Deutonymphal stage	2.9±0.350	2.36±0.21	
Total immature	7.45±0.669	6.38±1.23	
Life cycle	9.25±0.669	8.19±1.59	

During adulthood, longevity of *A. Cydnodactylon* female and male took an average of 21.45 and 18.15 days, respectively. The life span was 30.7 and 26.34 days, for female and male, respectively.

Regarding to the fecundity, the number of deposited eggs per female was 31.2 eggs, with a daily rate of 1.79 eggs (Table 2).

Periods and Fecundity	Mean±SE. days	
	Female	Male
Pre-oviposition	1.35±0.242	-
Oviposition	17.35±1.06	-
Post- oviposition	2.75±0.540	-
Longevity	21.45±1.04	18.15±0.629
Life span	30.7±1.31	26.34±1.28
Fecundity		
* No. of eggs/ female	31.2±2.098	-
* No. of eggs/ female/day	1.79±0.603	-

Table (2): Duration of longevity, life span and fecundity of *A. cydnodactylon*, fed on crawlers of *B. tabaci* at 25 ± 2 °C and 70 ± 5 % R.H.

Food consumption of A. cydnodactylon:

The consumption rate increased through the developmental stages, respectively (Table 3). The larval stage developed to the protonymphal stage without feeding. The female longevity of *A. cydnodactylon* was 21.45 days while that of male was 18.15 days. The consumption of *B. tabaci* nymphs

was more by female predator (274.1 nymphs) than male (205.34 nymphs). The maximum means for total food consumption of the female predator was recorded during the oviposition period, it consumes an average of (242.61 prey). Finally, the average number of devoured victim during life span were 303.8 and 212.65 preys for female and male, respectively.

Table (3): Total prey consumption of different life stages of A. cydnodactylon fed on crawlers of B. tabaci at 25 ± 2 °C and 70 ± 5 % R.H.

Consumption rate	+Mean ± SE. days	
	Female	Male
Larva	0.0 ± 0.0	0.0 ± 0.0
Protonymph	11.6±1.174	8.3±0951
Deutonymph	18.1±1.197	9.1±1.246
Life cycle	29.7±1.197	17.3±1.629
Longevity	274.1±13.739	205.34±11.28
Life span	303.8±14.32	212.65±13.74

4. Discussion

Little previous study has been made concerning the predation of this species; therefore I could not compare the results with previous published studies. However, there are numerous investigations on other phytoseiid species, revealing the effect *B. tabaci* on the biological aspects of *A. cydnodactylon*.

Results demonstrate that local population of *A. cydnodactylon* is able to feed and complete developmental stages when fed on the crawlers of *B. tabaci*.

The present results agree with, **El Banhawy**, *et al.* (2001). They reported that, *A. cydnodactylon* able to develop and reproduce when fed on other different prey species such as, nymphal stages of *Bemisiatabaci*, *Thripstabaci* (Lindeman) and *Tetranychus urticae*. They added, consumption rate of *A. cydnodactylon* increased as the development progressed. Also **Nomikou** *et al.* (2001) mentioned

that the phytoseiid mite, *Amblyseius swirskii* (Athias-Henriot), feed on eggs and crawlers of *B. tabaci* and develops well on this prey.

The present study showed that temperature affects the feeding capacity of all life stages of *A. cydnodactylon* except the larval stage, where it developed to the protonymphal stage without feeding. Non feeding larval behavior may be a mechanism to avoid sibling cannibalism. Similar findings have been reported for other phytoseiid species by **Zhang** et al. 1998; El-Banhawy et al. 2000; Chittenden & Saito, 2001; Kouhjani et al., 2009 and Fatemeh et al. 2011.

During immature stages of *A. cydnodactylon*, food consumption rate increased at the experimental temperature (25°C). **Metwally** *et al.*, **2005** and **Fatemeh** *et al.*, **2011** showed that the food consumption increased from 20 to 25°C. They added,

it could be concluded that the optimal temperature for predation of this predator was about 25°C.

The present data showed that the predator females during oviposition consumed a significantly higher number of prey, suggesting that females need extra food for egg production during this period. This information is in agreement with other findings of Tanigoshi, et al., 2001; Saha, et al., 2001; Kouhjani et al., 2009 and Omar and Mesbah 2013.

Although, I observed the same trend in all experiments, the obtained values were different because of different prey and predator species were used in the experiments. Furthermore, several other factors, such as relative humidity, photoperiod, presence of pollen, and the type of experimental arena may also affect a predator's feeding (Fernando and Hassell, 1980).

The results from the current study would help us to gain a better insight into the efficiency and practical application techniques of a predator in biological control programs of spider mites. According to the findings, *A. cydnodactylon* could be a beneficial biocontrol agent for *B. tabaci* in both greenhouses and field when temperature is 25°C; however, to optimize results, additional experiments should be performed. **Gerling, et al. (2001)** reported that reviews current efforts in biological control of *B. tabaci* in greenhouse and field crops, and highlights research gaps and directions deserving further development.

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References

- Abd-Rabou, S. and A.M. Simmons (2010): Survey of Reproductive Host Plants of *Bemisia* tabaci (Hemiptera: Aleyrodidae) in Egypt, Including New Host Records. Entomological News 121(5):456-465.
- 2. Brown, J.K. (2001): The Molecular Epidemiology of Begomoviruses, In J.A. Khan and J. Dykstra [eds.] Trends in Plant Virology. The Haworth Press, Inc., NY. pp. 279-316.
- 3. Chittenden, A.R. and Y. Saito, 2001. Why are there feeding and non-feeding larvae in phytoseiid mites (Acari: Phytoseiidae). J. Ethol. 19: 55–62.
- 4. De Barro, PJ. (1995): *Bemisia tabaci* biotype B: a review of its biology; distribution and control. CSIRO Technical Paper 1995:36:58.
- 5. El-Banhawy, E.M.; S.A.A. Amer and S.A. Saber (2000): Development and reproduction of the

- predacious mite, *Amblyseius cydnodactylon* on different prey species; effect of plant leaf texture on the behavior and reproduction of the predator. Z. Pflanzenkr. Pflanzenschutz, 107:218–224.
- El-Banhawy; E.M.; S.M. Hafez and S.A. Saber (2001): Response of Amblyseius cydnodactylon (Phytoseiidae) to increasing prey density of Tetranychus urticae (Tetranychidae) in absence or presence of nymphs of Bemisia tabaci (Homoptera) in Egypt. International Journal of Acarology, Vol. 27, (3): 241-244.
- 7. El-Sharkawy H., Mersal, R. And El Muttardy, F.(2013):Tomato yellow leaf curl virus :Efficiency of acquisition, retention and transmission by *Bemisia tabeci* (Gennadius),in Libya. J.Product.&Dev.,18(3):437-445
- 8. Fatemeh G.; F. Yaghoub and K. Karim, (2011): Effect of temperature on prey consumption of *Typhlodromus bagdasarjani* (Acari: phytoseiidae) on *Tetranychus urticae* (acari: Tetranychidae). International Journal of Acarology. Vol. 37,(6):556–560.
- 9. Fernando, M. H. J. P. and M. P. Hassell, (1980): Predator-prey responses in an acarine system. Res. Popul. Ecol. 22: 301–322.
- 10. Gennadius, P. (1889): Disease of tobacco plantations in Trikonia: the aleurodid of tobacco. Ellenike Georgia 5: 1-3.
- 11. Gerling, D. (Ed.), (1990): Whiteflies: Their Bionomics, Pest Status and Management. Athenaeum Press, New Castle, UK:pp 348.
- 12. Gerling, D.; O. Alomar and J. Arno (2001): Biological control of *Bemisia tabaci* using predators and parasitoids. Crop Protection 20:779–799.
- 13. Heinz, K. (1996): Predators and parasitoids as of *Bemisia tabaci* in biological control agents greenhouse. In: Gerling D, Mayer RT, Editors. taxonomy, Biology, Damage Control and Management. Intercept Ltd. pp. 439-449.
- 14. Hodges, G.S. and G.A. Evans (2005): An identification guide to the whiteflies (Hemiptera: Aleyrodidae) of the southeastern united states. Florida Entomologist, 88(4): 518-534.
- 15. Jones, D.R. (2003): Plant viruses transmitted by whiteflies Eur. J. Plant Pathol., 109: 195–219.
- 16. Kouhjani G. M.; Y. Fathipour and K. Kamali, 2009. The effect of temperature on the functional response and prey consumption of *Phytoseius plumifer* (Acari: Phytoseiidae) on the two-spotted spider mite. Acarina 17(2): 231–237.
- 17. Metwally, A.M.; B.A. Abou-Awad and M.M.A. Al-Azzazy, 2005. Life table and prey consumption of the predatory mite *Neoseiulus cydnodactylon* Shehata and Zaher (Acari:

- Phytoseiidae) with three mite species as prey. Z. Pflanzenk, Pflanzen.112: 276–286.
- 18. Mughra, R.B.; S.S. Liu and X. Zhou (2008): Tomato yellow leaf curl virus and tomato leaf curl Taiwan virus invade southeast coast of China J. Phytopathol., 156:217–221.
- Nomikou, M.; A. Janssen; R. Schraag and M.W. Sabelis (2001): Phytoseiid predators as potential biological control agents for *Bemisia tabaci*. Experimental and Applied Acarology, 25: 271-291.
- 20. Omar N.A. and A.E. Mesbah (2013): The effect of temperature on development, fecundity and food consumption of *Amblyseius cydnodactylon* Shehata & Zaher (Acari: Phytoseiidae). Acarines, Egypt. J. Agric. Res. Vol. 13(1):45-55.
- 21. Palumbo, J.C.; A.R. Horowitzand N. Prabhaker (2001): Insecticidal control and resistance management for *Bemisia tabaci*. Experimental & Applied Acarology, Vol. 25, 4: 271-291.
- 22. Qiu, B.; S.A. Coats and S.X. Ren (2007): Phylogenetic relationships of native and introduced *Bemisia tabaci* (Hemiptera: Aleyrodidae) from China and India based on mtCOI DNA sequencing and host plant comparisons Prog. Nat. Sci., 17: 645–654.
- 23. Qiu, B.; Y. Chen; L. Liu; W. Peng; X. Li; M.Z. Ahmed; V. Mathur; D. Yuzhou and S. Ren

- (2009): Identification of three major *Bemisia tabaci* biotypes in China base on morphological and DNA polymorphisms. Progress in Natural Science 19: 713–718.
- 24. Saha K.; A.K. Somchoudhury; P.K. Sarkar and S.K. Gupta (2001): Effect of temperature on the rate of development, fecundity, longevity, sex ratio and mortality of *Amblyseius coccosocius* Ghai and Menon (Acari: Phytoseiidae), an important biocontrol agent against tea red spider mite in India. Acarology: Proceedings of the 10th International Congress: 470-472.
- 25. Snedecor, G.W. and G.W. Cochran (1982): Statistical Methods. Iowa State Uinv., Press, 7 Edition Ames, USA.
- 26. SPSS, (2006): SPSS User's Guide Statistics. Version 10. Copyright SPSS Inc., USA.
- Tanigoshi, L.K.; S.C. Ho Gerling, D.; O. Alomar and J. Arno (2001): Biological control of *Bemisia tabaci* using predators and parasitoids. Crop Protection 20: 779–799.
- 28. Zhang, Z.Q.; L. Jianzhen and L. Qiaoyun, 1998. Predation of *Amblyseius longispinosus* (Acari: Phytoseiidae) on *Aponychus corpuzae* (Acari: Tetranychidae). Systematic and Applied Acarology 3: 53-58.

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